Three New Thiophene Acetylenes from *Rhaponticum uniflorum* (L.) DC.

by Hai-Li Liu and Yue-Wei Guo*

State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Zu Chong Zhi Rd. 555, Zhangjiang Hi-Tech Park, Shanghai 201203, P. R. China (phone: +86-21-50805813; e-mail: ywguo@mail.shcnc.ac.cn)

Three new thiophene-acetylenes, 7-chloroarctinone-b (1), rhapontiynethiophenes A (2) and B (3), along with arctinone-b (4) were isolated from the MeOH extract of the roots of *Rhaponticum uniflorum* (L.) DC. The structures of the new compounds were elucidated by detailed spectroscopic data analysis and by comparison with known compounds.

Introduction. – Thiophene-acetylenes (=ethynylthiophenes) represent a unique class of natural products that often exhibit a wide variety of biological activities ranging from antitumor [1], antiviral [2], anti-HIV [3], antifungal [4] to insecticidal activities [5][6]. Thiophene-acetylenes are typically found in the plant family Asteraceae [7][8], but they have also been reported in fungi [9].

Rhaponticum uniflorum (L.) DC. (family Asteraceae) is a perennial herbaceous plant widely distributed in the northern part of China. The roots of *R. uniflorum* (commercial name: *Qizhou Loulu*) have been used as folk medicine for the treatment of fever and intoxications [10] in China. Previous phytochemical studies on the title plant have resulted in the isolation of phytoecdysteroids [11], triterpenes [12], diterpenes [13] and thiophene-acetylenes [14–16]. In the course of our search for bioactive secondary metabolites from the Chinese fauna and flora [17–19], we have reinvestigated this medicinal plant. Chemical investigation of three new thiophene-acetylenes, 7-chloroarctinone-b (1), rhapontiynethiophenes A and B (2 and 3, resp.), together with arctinone-b (4), which has previously been isolated from the same plant [14]. Here, we describe the isolation and structure elucidation of the new compounds 1-3.

Results and Discussion. – The powdered roots of *R. uniflorum* collected from Chifeng City, Inner Mongolia Autonomous Region, China, were extracted exhaustively with MeOH. The MeOH extract was then partitioned between H_2O and AcOEt, and H_2O and BuOH, respectively. The AcOEt-soluble portion was repeatedly subjected to silica-gel and *Sephadex LH-20* column chromatography, followed by reversed-phase (RP) HPLC purification, resulting in the isolation of four thiophene-acetylenes, of which three (7-chloroarctinone-b (1), and rhapontiynethiophenes A and B (2 and 3, resp.)) were new thiophene-acetylenes (*Figure*). The known compound was identified

^{© 2008} Verlag Helvetica Chimica Acta AG, Zürich



Figure. Chemical structures of compounds 1-5¹)

as arctinone-b (4) [14] by analysis of its NMR spectra and by comparison with the literature data.

All of the new compounds exhibited very similar spectroscopic properties to cooccurring arctinone-b (4). Their UV spectra showed strong absorption maxima around 260 nm and, in the case of 1 and 2, 371 or 340 nm, respectively, which was indicative for the presence of one or two thiophene rings [20]. The presence of a thiophene ring was further supported by typical IR absorptions at $\tilde{\nu}_{max}$ ca. 1400 and ca. 800 cm⁻¹ [21]. The NMR spectra of 1–3 (*Tables 1* and 2) show strong structural analogies with known thiophene-acetylenes, which possess one or two 2,5-disubstituted thiophene moieties (confirmed by comparison of the ¹H- and ¹³C-NMR data with those of model compounds 4 [14] and 5 [22]). In fact, compounds 1 and 2 like co-occurring 4 possess a common 2,2'-dithiophene-5'-(prop-1-ynyl) partial structure and differ from each other only by the substituents at C(5), while compound 3 differs from the model compound 5, which was previously obtained from *Leuzea carthamoides* [22], only by the substituent at C(2).

Compound **1**, 7-chloroarctinone-b²), was obtained as a yellow powder. The molecular formula, $C_{13}H_9ClOS_2$, consistent with nine degrees of unsaturation, was determined by HR-EI-MS (*m*/*z* 279.9775, *M*⁺; calc. 279.9783). Analysis of 1D- and 2D-(¹H, ¹H-COSY, HMQC, and HMBC) NMR spectra (see *Table 1*) readily allowed us to recognize a typical 2,2'-dithiophene moiety with two substituents at C(5) and C(5'), respectively. Moreover, the characteristic ¹³C-NMR chemical shifts at δ (C) 92.9 (*s*), 72.5 (*s*), and 4.8 (*q*) indicated the presence of a prop-1-ynyl group in **1**.

The above-mentioned structural features of **1** were strongly reminiscent of those of co-occurring arctinone-b (**4**) [14]. Careful comparison of the NMR data of **1** with those of **4** revealed that the only difference between **1** and **4** exists at C(5), where the Ac function in **4** is replaced by a 7-chloro group in **1**, in agreement with a difference of 34 mass units. This conclusion was further supported by the observation of a downfield ¹H-NMR signal at δ (H) 4.55 (*s*, 2 H) in **1**, accompanying the disappearance of the Me

¹) Arbitrary numbering, see also in the *Figure*.

²) For systematic names, see Exper. Part.

| Position ¹) | 1 | | 2 | |
|-------------------------|----------------------|---------------------------|----------------------|----------------|
| | $\delta(\mathrm{H})$ | $\delta(\mathrm{C})^{c})$ | $\delta(\mathrm{H})$ | $\delta(C)^c)$ |
| C(2) | - | 146.5(s) | - | 135.8 (s) |
| H-C(3) | 7.15 (d, J = 3.9) | 124.3(d) | 6.90 (d, J = 3.9) | 123.0(d) |
| H-C(4) | 7.67 $(d, J = 3.9)$ | 134.0(d) | 6.97 (d, J = 3.9) | 126.9(d) |
| C(5) | _ | 138.6(s) | _ | 129.3(s) |
| C(6) | _ | 183.8(s) | _ | - |
| $CH_{2}(7)$ | 4.55(s) | 45.1(t) | _ | - |
| C(2') | _ | 135.5(s) | _ | 136.5(s) |
| H-C(3') | 7.16 (d, J = 3.9) | 125.7(d) | 6.91 (d, J = 3.9) | 122.7(d) |
| H-C(4') | 7.04 (d, J = 3.9) | 132.1(d) | 6.82(d, J = 3.9) | 131.7(d) |
| C(5') | _ | 126.0(s) | _ | 124.0(s) |
| C(6') | _ | 72.5(s) | _ | 72.6(s) |
| C(7') | _ | 92.9(s) | _ | 91.5 (s) |
| Me(8') | 2.10 (s) | 4.8(q) | 2.09 (s) | 4.7 (q) |

^a) *Bruker DRX-400* spectrometer, assignments made by HMQC and HMBC experiments. ^b) In CDCl₃, chemical shift δ [ppm] referred to the solvent signal (δ (H) 7.26, δ (C) 77.0); *J* in Hz. ^c) Multiplicities by DEPT sequence.

Table 2. ¹H- and ¹³C-NMR Data^a)^b) for **3** and ¹³C-NMR Data^b) for **5**

| Position ¹) | 3 | | 5 [22] |
|-------------------------|--------------------------|------------------|---------------|
| | $\delta(\mathrm{H})$ | $\delta(C)^{c})$ | $\delta(C)$ |
| C(2) | _ | 121.4(s) | 121.9 (s) |
| H-C(3) | 7.16(d, J = 3.9) | 137.1(d) | 135.1(d) |
| H-C(4) | 6.78(d, J = 3.9) | 125.7(d) | 125.2(d) |
| C(5) | _ | 153.1(s) | 151.4(s) |
| H-C(6) | 4.85 (dd, J = 3.6, 7.5) | 72.4(d) | 71.9(d) |
| $H_a - C(7)$ | 3.68 (dd, J = 3.6, 11.7) | 68.7(t) | 68.3(t) |
| $H_{\rm b} - C(7)$ | 3.58 (dd, J = 7.5, 11.7) | | () |
| C(1') | _ | 69.8(s) | 74.2(s) |
| C(2') | _ | 79.4(s) | 78.5(s) |
| C(3') | _ | 59.5(s) | 72.5(s) |
| C(4') | _ | 65.5(s) | 83.7 (s) |
| C(5') | _ | 70.9(s) | 110.6(d) |
| C(6') | _ | 81.4(s) | 145.4(d) |
| Me(7') | 1.95 (s) | 4.5 (q) | 19.0 (q) |

^a) *Bruker DRX-400* spectrometer, assignments made by HMQC and HMBC experiments. ^b) In CD₃OD, chemical shift δ [ppm] referred to the solvent signal (δ (H) 3.30, δ (C) 49.0); *J* in Hz. ^c) Multiplicity by DEPT sequence.

signal at $\delta(H)$ 2.10 (s, 3 H) in **4**. Moreover, the presence of a Cl-atom was clearly indicated by the M^+ peaks at m/z 279.9775 and 281.9748 with intensities of 1:0.44 in the HR-EI-MS. Accordingly, the structure of **1** was determined to be 7-chloroarctinone-b.

Table 1. ¹H- and ¹³C-NMR Data^a)^b) for **1** and **2**

Rhapontiynethiophene A (2), a yellow oil, had the molecular formula $C_{11}H_7ClS_2$ which is 8 mass units less than for 4. Comparison of the NMR data of 2 and 4 revealed that the only difference was located at C(5) where the Ac group in 4 was replaced by a Cl-substituent in 2, while the rest of the structure of 2 was identical to 4. Consequently, the structure of compound 2 was established as 5-chloro-5'-(prop-1-ynyl)-2,2'-bithiophene.

Rhapontiynethiophene B (3), an optically active yellow powder, showed the molecular formula of $C_{13}H_{10}O_2S$, as established by HR-EI-MS (m/z 230.0399 (M^+); calc. 230.0402), indicating nine degrees of unsaturation. The typical ¹H-NMR signals at δ (H) 7.16 (d, J = 3.9, H-C(3)) and 6.78 (d, J = 3.9, H-C(4)), and ¹³C-NMR signals at $\delta(C)$ 59–82 ppm (*Table 2*) indicated that **3** is also a thiophene-acetylene. In addition, the presence of a 1,2-diol partial structure was revealed by the explicit ¹H,¹H-COSY correlation from H-C(6) to $CH_2(7)$, together with the diagnostic ¹³C-NMR chemical shifts at $\delta(C)$ 72.4 (C(6)) and 68.7 (C(7)). A literature survey showed that the NMR data of 3 (Table 2) were almost identical to those of the model compound 5 (1-{5-[(5E)-hept-5-ene-1,3-diynyl]thiophen-2-yl}ethane-1,2-diol) [22] with the only exception at the terminus of the side chain at $C(2)^1$, where the C=C bond in 5 was replaced by a C=C bond (δ (C) 70.9 and 81.4) in **3**, in agreement with a difference of 2 mass units. Detailed analysis of HMQC and HMBC spectra of 3 allowed the unambiguous assignment of the planar structure of **3**. Especially, HMBC correlations from Me(7') $(\delta(H) 1.95)$ to C(5') and C(6') confirmed this conclusion (Figure). Rhapontiynethiophene B (3) is the 5',6'-dehydro derivative of 5.

Compound **3** showed a negative optical rotation $([\alpha]_D^{22} = -13.1 (c = 0.38, \text{MeOH}))$, though the related compound **5** was reported to be a racemate. The absolute configuration at C(6) of **3** remains undefined due to the limited amounts of **3** obtained from the plant.

Chlorinated secondary metabolites are rather rare from terrestrial sources. In this context, it cannot be definitively ruled out that compounds **1** and **2** are artifacts formed during the isolation procedure. However, it must be pointed out that several chlorinated thiophene-acetylenes have been previously reported from the family Asteraceae [23-29]. It is noteworthy that several of them were obtained without the use of chlorinated solvents during the extraction and isolation procedures [27-29].

This research work was financially supported by National '863' Project (No. 2006AA09Z412), the *Natural Science Foundation of China* (Nos. 30730108, 20721003), CAS Key Project (grant KSCX2-YW-R-18) and STCSM Projects (No. 054307062 and 06DZ22028). We are indebted to Associate Prof. *J.-G. Shen* for identification of the plant.

Experimental Part

General. Column chromatography (CC): commercial silica gel (*Qing Dao Hai Yang Chemical Group* Co., 100–200 and 200–300 mesh) or Sephadex LH-20 (Amersham Biosciences). TLC: Precoated silicagel plates (Yan Tai Zi Fu Chemical Group Co., G60 F-254) were used for anal. TLC. Reversed-phase (RP) HPLC: Agilent 1100 series liquid chromatography using a VWD G1314A detector at 210 nm. A semi-preparative ODS-HG-5 (5 µm, 9.4 mm (i.d.) × 25 cm) was employed for the purification. Optical rotation: Perkin-Elmer 341 polarimeter. UV Spectra: Varian Cary-300-Bio spectrophotometer; λ_{max} (log ε) in nm. IR Spectra: Nicolet Magna-FT-IR 750 spectometer; \tilde{v}_{max} in cm⁻¹. ¹H- and ¹³C-NMR Spectra: Bruker DRX-400 (400 MHz for ¹H, and 100 MHz for ¹³C) spectrometer; chemical shift δ in ppm, with the solvent signal in CDCl₃ (δ (H) 7.26, δ (C) 77.0) or in CD₃OD (δ (H) 3.30; δ (C) 49.0) as an internal standard, coupling constant *J* in Hz; assignments supported by HMQC and HMBC experiments. EI-MS and HR-EI-MS: *Carlo-Erba TRIO 2000 VG* and *Kratos MS50* mass spectrometers, in *m*/*z*.

Plant Material. The plants were collected at Chifeng City, Inner Mongolia Autonomous Region, People's Republic of China, in March 2007 and identified as *R. uniflorum* by Associate Prof. *J.-G. Shen* of the Shanghai Institute of Materia Medica, Chinese Academy of Sciences. A voucher specimen (No. 0703P-40) is available for inspection at the Herbarium of Shanghai Institute of Materia Medica, CAS.

Extraction and Isolation. The powdered dry roots (5 kg) of *R. uniflorum* were exhaustively extracted with MeOH (101×3 ; 3 d each time) at r.t., and the MeOH extract was concentrated *in vacuo* to give a residue (210 g), which was suspended in H₂O (1000 ml), and then sequentially extracted (3×11) with AcOEt and BuOH, to afford an AcOEt-soluble extract (40 g) and an BuOH-souble extract (20 g), resp. The AcOEt extract was fractionated by gradient silica-gel column chromatography (0-100% AcOEt in light petroleum ether (PE), successively) yielding three fractions showing interesting green TLC spots after spraying with H₂SO₄ (R_f 0.55 (PE), 0.60 and 0.22 (PE/Me₂CO 7:3), resp.). These fractions were further purified by repeated silica-gel CC (PE/Et₂O, PE/acetone, and CHCl₃/MeOH), followed by *Sephadex LH-20* CC (PE/CHCl₃/MeOH 2:1:1 and CHCl₃/MeOH 1:1), yielding pure compounds **2** (4.5 mg) and **3** (2.0 mg), resp., and a mixture, which was separated into other two pure compounds **1** (22.8 mg) and **4** (8.2 mg) by RP-HPLC (semi-prep. *ODS-HG-5* (5 µm, 9.4 mm (i.d.) × 25 cm), MeOH/ H₂O (80%), 2.0 ml/min).

7-Chloroarctinone-b (=2-Chloro-1-[5'-(prop-1-ynyl)](2,2'-bithiophen]-5-yl]ethanone; **1**). Yellow powder. UV (MeOH): 371 (4.45), 262 (4.09), 220 (3.92). IR (KBr): 2210, 1640, 1510, 1455, 1420, 1351, 1271, 1250, 1024, 800. ¹H- and ¹³C-NMR: *Table 1*. EI-MS: 282 (30, $[M+2]^+$), 280 (72, M^+). HR-EI-MS: 279.9775 (M^+ , C₁₃H₉ClOS⁺₂; calc. 279.9783).

Rhapontiynethiophene A (=5-*Chloro-5'-(prop-1-ynyl)-2,2'-bithiophene*; **2**). Yellow oil. UV (MeOH): 340 (4.40), 260 (4.11), 241 (3.74). IR (KBr): 2211, 1439, 810, 797. ¹H- and ¹³C-NMR: *Table 1*. EI-MS: 240 (40, $[M+2]^+$), 238 (100, M^+). HR-EI-MS: 237.9681 (M^+ , C₁₁H₇ClS₂⁺; calc. 237.9678).

Rhapontiynethiophene B (=(1R*)-1-[5-(Hepta-I,3,5-triynyl)thiophen-2-yl]ethane-I,2-diol; **3**). Yellow powder. $[a]_{D}^{2D} = -13.1 (c = 0.38, MeOH). UV (MeOH): 269 (4.41), 253 (4.50), 211 (4.44). IR (KBr): 3437, 2214, 2173, 1383, 1086, 1040, 806. ¹H- and ¹³C-NMR:$ *Table 2*. EI-MS: 230 (32,*M* $⁺), 199 (100, <math>[M - CH_2OH]^+$). HR-EI-MS: 230.0399 (*M*⁺, C₁₃H₁₀O₂S⁺; calc. 230.0402).

REFERENCES

- [1] R. Ebermann, G. Alth, M. Kreitner, A. Kubin, J. Photochem. Photobiol., B 1996, 36, 95.
- [2] R. J. Marles, J. B. Hudson, E. A. Graham, C. Soucy-Breau, P. Morand, R. L. Compadre, C. M. Compadre, G. H. N. Towers, J. T. Arnason, *Photochem. Photobiol.* **1992**, *56*, 479.
- [3] J. B. Hudson, L. Harris, A. Teeple, G. H. N. Towers, Antiviral Res. 1993, 20, 33.
- [4] D. Mares, M. P. Fasulo, A. Bruni, J. Med. Vet. Mycol. 1990, 28, 469.
- [5] M. Nivsarkar, G. P. Kumar, M. Laloraya, M. M. Laloraya, Arch. Insect Biochem. Physiol. 1991, 16, 249.
- [6] A. Sharma, H. C. Goel, Indian J. Exp. Biol. 1994, 32, 745.
- [7] F. Bohlmann, T. Burkhardt, C. Zdero, in 'Naturally Occurring Acetylenes', Academic Press, London and New York, 1973.
- [8] J. B. Harborne, in 'Phytochemical Methods', 2nd edn., Chapman and Hall, New York, 1984, p. 171.
- [9] E. R. H. Jones, C. M. Piggin, V. Thaller, J. L. Turner, J. Chem. Res. 1977, 68.
- [10] Jiangsu New Medicinal College, in 'The Encyclopaedia of Traditional Chinese Medicine', Shanghai Science and Technology Press, Shanghai, 1985, p. 2348.
- [11] X.-F. Jiang, X. Li, Chin. Tradit. Herb. Drugs 1997, 28, 262.
- [12] Y.-H. Zhang, J.-K. Cheng, L. Yang, D.-L. Cheng, J. Chin. Chem. Soc. 2002, 49, 117.
- [13] B. Liu, R.-B. Shi, G.-Z. Tu, W. Wang, C.-M. Yang, Beijing Zhongyiyao Daxue Xuebao, 2004, 27, 58.
- [14] H.-X. Wei, W.-Y. Gao, Y.-J. Tian, Y.-K. Guan, M.-H. Huang, D.-L. Cheng, *Pharmazie* 1997, 52, 245.
- [15] M.-S. Liu, X. Li, S.-X. Wang, Chin. J. Med. Chem. 1996, 6, 121.

- [16] D.-A. Guo, Z.-C. Lou, C.-Y. Gao, D. Wang, H.-Y. Zhang, Chin. Tradit. Herb. Drugs 1992, 23, 178.
- [17] Z.-Y. Li, P. Chen, H.-G. Xu, Y.-M. Yang, S.-Y. Peng, Z.-Z. Zhao, Y.-W. Guo, Org. Lett. 2007, 9, 477.
- [18] W. Zhang, Y.-W. Guo, K. Krohn, Chem. Eur. J. 2006, 12, 5122.
- [19] J.-D. Wang, Z.-Y. Li, W.-S. Xiang, Y.-W. Guo, Helv. Chim. Acta 2006, 89, 1367.
- [20] J. W. Sease, L. Zechmeister, J. Am. Chem. Soc. 1947, 69, 270.
- [21] J. S. Serensen, T. Mortensen, N. A. Serensen, Acta Chem. Scand. 1964, 18, 2182.
- [22] V. Chobot, V. Buchta, H. Jahodářová, M. Pour, L. Opletal, L. Jahodář, P. Harant, Fitoterapia 2003, 74, 288.
- [23] K. L. Stevens, S. C. Witt, C. E. Turner, Biochem. Syst. Ecol. 1990, 18, 229.
- [24] Y.-Q. Tian, X.-Y. Wei, H.-H. Xu, J. Nat. Prod. 2006, 69, 1241.
- [25] Y. Liu, M. Ye, H.-Z. Guo, Y.-Y. Zhao, D.-A. Guo, J. Asian Nat. Prod. Res. 2002, 4, 175.
- [26] N. Fokialakis, C. L. Cantrell, S. O. Duke, A. L. Skaltsounis, D. E. Wedge, J. Agric. Food Chem. 2006, 54, 1651.
- [27] J. Lam, L. P. Christensen, T. Thomasen, Phytochemistry 1991, 30, 1157.
- [28] C. Zdero, F. Bohlmann, L. Haegi, R. M. King, Phytochemistry 1988, 27, 865.
- [29] F. Bohlmann, N. Borthakur, H. Robinnson, R. M. King, Phytochemistry 1982, 21, 1795.

Received July 20, 2007